



## 25 **RESUMO**

26 Parasitas gastrointestinais são freqüentemente encontrados em animais domésticos, com  
27 importante papel na saúde pública e animal. Este estudo teve como objetivo verificar a  
28 ocorrência de parasitas gastroentéricos em cães e gatos de Mineiros, Brasil, e seu impacto na  
29 saúde pública. No total, 103 amostras fecais (93 de cães e 10 de gatos) foram coletadas de abril  
30 de 2017 a julho de 2018 por defecação espontânea e processadas para a busca de ovos e oocistos  
31 pela técnica de Willis (adaptado). *Toxocara* spp. foi o parasita mais frequente identificado em  
32 cães, com 34,41% (32/93 amostras), seguido de *Ancylostoma* spp. (11,83%, 11/93 amostras) e  
33 *Isospora* spp. (1,07%, 1/93 amostras). Amostras de fezes de gatos apresentaram resultado  
34 semelhante, sendo *Toxocara* spp. o parasita mais frequente (40%, 4/10 amostras), seguido por  
35 *Ancylostoma* spp. (20%, 2/10 amostras) e *Isospora* spp. (20%, 2/10 amostras). Animais jovens  
36 foram 10% mais positivos para parasitas em comparação com animais idosos e adultos (odds  
37 ratio = 1,1), assim como animais de rua 20% mais parasitados (odds ratio = 1, 2). *Toxocara* spp.  
38 e *Ancylostoma* spp. São relatados como os parasitas mais comuns encontrados em cães e gatos  
39 no mundo, causando danos digestivos, inclusive fatais, e com grande importância na saúde  
40 pública. Programas eficazes de controle de parasitas animais, além da importância da educação  
41 em saúde, são necessidades óbvias de acordo com os dados do estudo.

42 **Palavras-chave:** Gastrointestinal.. Helmintos. Parasitologia. Protozoário. Zoonoses.

43

44

## **INTRODUCTION**

45 Gastrointestinal parasites are frequently found in domestic dogs and cats, playing an  
46 important role in animal and public health. Initiating in the animals, development delays and  
47 severe gastroenteritis, which may progress to death. In humans, they can cause cutaneous  
48 disorders, allergies, diarrhea, weight losses and dehydration (ZAJAC & CONBOY, 2012).

49 The occurrence of these endoparasites is mainly due to the exacerbated flow of animals  
50 in public places, since feces excreted by infected animals contaminate the environment,  
51 contributing to the infection of new hosts, whether animals or humans (SANTARÉM et al.,  
52 2004). Infection can occur either through vertical or horizontal transmission through  
53 contaminated food and water in most species, also by the percutaneous route (BOWMAN,  
54 2014).

55 Bowman (2014) and Sharif et al. (2007) described *Toxocara* spp. and *Ancylostoma* spp.,  
56 as the most common canine and feline parasites in regions with a tropical and subtropical  
57 climate. They are of great importance in public health, causing visceral and cutaneous *larva*  
58 *migrans* in humans. Since susceptible humans and animals are closely linked in everyday life,  
59 it is important to know their levels of occurrence.

60 The study aims to identify the frequency of gastrointestinal parasites in dogs and cats  
61 from Mineiros, Goiás, to determine which helminths and protozoans are most frequently.

62

63

### MATERIALS AND METHODS

64 From April 2017 to July 2018 were analyzed canine and feline fecal samples of 103  
65 animals from Mineiros, Goiás, Brazil. Samples were collected by spontaneous defecation and  
66 immediately processed to search for helminth eggs and protozoan oocysts. The sample size  
67 calculation was calculated following the formula:

68

$$69 \quad n = \left( \frac{Z_{(1-\alpha)} * \sqrt{p * (1 - p)}}{d} \right)^2 = \left( \frac{1.96 * \sqrt{0.76 * 0.24}}{0,1} \right)^2 \cong 71 \text{ (minimum)}$$

70

71 Where  $n$  is the sample number;  $Z_{(1-\alpha)}$  is the  $Z$  value of the standard normal curve for the  
72 degree of confidence ( $1-\alpha$ );  $p$  is the estimated proportion of expected results; and  $d$  is the  
73 desired precision. The value of  $Z_{(1-\alpha)}$  was set at 1.96 (for a 95% confidence level). The  
74 estimated proportion  $p$  was established using data on the maximum gastrointestinal parasites  
75 occurrence in dogs from miwestern of Brazil (RAMOS et al., 2015), same region of study area.  
76 Finally, for  $d$  the value of 0.1 for accuracy at the 10% level

77 After collected, the samples were processed according to the protocol established by  
78 Willis (1921), described and adapted by Hoffmann (1987), where they were mixed with sodium

79 chloride hypersaturated solution (NaCl - 35%), filtered in sieve and gauze to remove dirt and  
80 subjected to spontaneous fluctuation of eggs and intestinal endoparasite oocysts by density  
81 difference. After this procedure, they were analyzed between slide and coverslip in optical  
82 microscope (Nikon Eclipse E200) with lugol addition to facilitate the observation of eggs by  
83 staining and eggs and oocysts were identified according to Zajac and Conboy (2012).

84 The level of occurrence will be presented in percentages, described by Bush et al. (1997),  
85 considering the number of positive animals compared to the number of samples collected. *Odds*  
86 *ratio* will be used to compare the risks of infection, considering the risk factors age and street  
87 access.

88

89

## RESULTS AND DISCUSSION

90 In total 93 fecal samples were collected from dogs and 10 fecal samples from cats  
91 (N=103), and 45 samples were positive to gastrointestinal parasites, and eggs of *Toxocara* spp.  
92 and *Ancylostoma* spp, and oocysts of *Isospora* spp, were identified.

93 Among 93 canine fecal samples, 41.94% (39 samples) were positive. *Toxocara* spp. was  
94 the most frequent parasite 31.41% (32 samples) of parasitism, followed by 11.83% (11 samples)  
95 for *Ancylostoma* spp. and 1.07% (1 sample) for *Isospora* spp. Mixed occurrences were detected  
96 in five samples, where *Toxocara* spp. and *Ancylostoma* spp. were identified. (Figure1).

97 Parasites were found in 60% (6 samples) of feline fecal samples, being frequencies 40%  
98 (4 samples) *Toxocara* spp., 20% (2 samples) *Ancylostoma* spp., and 20% (2 samples) *Isospora*  
99 spp.. Mixed infections were observed in two samples, being *Toxocara* spp. and *Ancylostoma*  
100 spp. in a sample, and *Toxocara* spp. and *Isospora* spp. in other.

101 Dogs and cats were classified as young (less than one year), adult (one to seven years)  
102 and elderly (older than seven years). Young animals were 10% more likely to be positive for  
103 parasites comparing to other ages (*odds ratio*=1.1). For the elderly, the ratio is 0.86, which is

104 14% less likely to become infected. In adults, the result is 0.68, that is, they have 32% less  
105 chances of infection.

106 About street access, animals with street access showed most chance to parasite infection,  
107 where the *odds ratio* was 1.2, 20% more likely to be parasitized. Animals without access to the  
108 street, presents 18% less chances of being infected with parasites (*odds ratio*=0.82).

109 Similar occurrences os infected animals by gastrointestinal parasites was observed in  
110 Ribeirão Preto (41.7%) (CAPUANO & ROCHA, 2006), Campo Grande (56.8%) (ARAÚJO et  
111 al., 1999), and Cuiabá (60.96% positive results for felines and 76% for canines) (RAMOS et  
112 al., 2013; RAMOS et al., 2015). However, many factors can interfere with the fluctuation of  
113 the occurrence, so there are very different percentages, depending on the location and the  
114 period.

115 Belonging to the Ascaridae family, the parasite of the genus *Toxocara* spp. is one of the  
116 largest nematodes that parasitize the small intestine of mammals, causing toxocarariasis in  
117 animals and visceral *larva migrans* (BOWMAN, 2014) which is an anthroponosis first  
118 described in 1907 in Cambridge in a study performed on dogs autopsies (LEIPER, 1907). Eggs  
119 eliminated through infected animals waste are very resistant to moisture and survive at  
120 temperatures from 10 to 45°C and can persist in the environment for five years (MARTINS,  
121 2019).

122 Woodruff et al. (1981) describe that desiccation and sunlight are factors that decrease the  
123 infectious capacity of the parasite. These factors are clearly observed in the areas of this study,  
124 which suffer from low humidity, high temperatures and the occurrence of fires, especially  
125 during dry season. During the rainy season, on the other hand, Cerrado region has very high  
126 humidity, so it is an environment that is advantageous during rain and disadvantageous during  
127 drought. However, it is noteworthy that the presence of wandering animals and urban areas that

128 provide microclimates for the development of eggs with the infecting larva keeps the  
129 occurrence of this parasite among animals (CAPUANO & ROCHA, 2006).

130 Eggs eliminated through feces containing L2 at ideal temperatures become infective after  
131 four weeks. Once ingested, L2 hatch in the small intestine, travel through the bloodstream to  
132 the lungs, where it becomes L3 and return through trachea to the intestine for its last two  
133 seedlings to adulthood. This form of infection occurs regularly in animals up to three months  
134 old (BOWMAN, 2014). Considering that the most frequent parasite was *Toxocara* spp. in the  
135 period and in the studied area, this justifies the greater risk of infection in younger animals,  
136 since the most abundant parasite affects this age group. Young animals still forming immune  
137 system, unable to respond to infections properly, suffer the most when parasitized, and can be  
138 infected by the transplacental or transmammary transmission. However, older animals can also  
139 be affected, especially when elderly or immunosuppressed (RAMÍREZ-BARRIOS et al.,  
140 2004).

141 Clinical signs are most commonly seen in puppies, in which adult worms begin to cause  
142 discomfort and increased abdominal volume, moans, opaque hair, vomiting and diarrhea  
143 (SHERDING & JOHNSON, 2008). Human infection occurs accidentally through the helminth  
144 in its still larval stage or from embryonated parasite eggs through ingestion when present in  
145 contaminated, raw, poorly sanitized or undercooked food. The parasite migrates through the  
146 body causing an inflammatory reaction. Clinical signs depend on the host immune response,  
147 ranging from asymptomatic to more severe cases (PAWLOWSKI, 2001).

148 *Ancylostoma* spp. cause cutaneous *larva migrans* (CLM) or ancylostomiasis, popularly  
149 known as the “hookworm infection”, is an anthrozoosis distributed mostly in tropical and  
150 subtropical countries, as well as the Midwest of Brazil (SANTARÉM et al., 2004). The  
151 *Ancylostoma* spp. genus has a high biotic potential (200-6000 eggs) and active larval infection  
152 capacity (L3), which can be found in the small intestine of dogs and cats. In addition, eggs can

153 hatch in just five days under optimal conditions and develop rapidly to L3 in tropical areas with  
154 sandy soils, where temperatures can be found between 25 ° C and 30 ° C (BOWMAN, 2014),  
155 climate completely compatible with the city of Mineiros in the Midwestern of Brazil.

156 CLM is generally related to people who have had contact with sand, especially children  
157 who, in addition to exposing the skin to possible infection, can also ingest eggs through soil  
158 contamination (NUNES et al., 2000). After skin penetration, the larvae migrate through the  
159 subcutaneous tissue, causing itching and rashes because of the inflammatory reaction.

160 In animals after percutaneous or oral infection, larvae can migrate through the  
161 bloodstream to the lungs, bronchi and trachea, where they become L4. They are then swallowed  
162 and move to the small intestine, where it transforms into adults (BOWMAN, 2014). Each worm  
163 as an adult extract about 0.1 ml of blood a day from its host, and in massive infections, animals  
164 quickly develop intense anemia. Parasitized animals suffer from weight loss, dehydration and  
165 may become depressed and may even die (RIBEIRO, 2004).

166 *Isospora* protozoa have a wide variety of hosts (TAYLOR et al., 2007). The most  
167 common clinical signs affecting infected animals are mild diarrhea, rarely presenting  
168 hemorrhage, and in more advanced cases, vomiting and dehydration (VASCONCELOS et al.,  
169 2008). The environment is contaminated through the excretion of feces from infected hosts  
170 containing unsporulated oocysts, where at appropriate temperature, humidity and oxygenation,  
171 they sporulate and become infective. After sporulation, the oocyst presents two sporocysts  
172 containing four sporozoites each (TAYLOR et al., 2007). Martins (2019) reported that ingestion  
173 of infected food, water or even rodents are the main means of infection to new hosts.

174 By passing through the digestive tract, sporulated oocysts release sporozoites into the  
175 intestinal lumen, which enter the enterocytes, where they begin their endogenous development.  
176 The parasite then begins its proliferative phase within the cells, asexually, through successive  
177 meiosis, where there is formation of schizonts containing merozoites. The invaded intestinal

178 cells rupture under pressure, releasing the merozoites, where the sexual phase begins, forming  
179 male and female gametes, thus forming the oocyst by the union of these gametes (MARTINS,  
180 2019). In mild infections, the intestinal mucosa suffers only a decrease in local absorption. On  
181 the other hand, in more severe infections, where schizonts develop deep in the mucosa, severe  
182 lumen destruction occurs, causing hemorrhage (TAYLOR et al., 2007).

183 In general, the most affected animals by isosporosis are the pups, which usually acquire  
184 the infection through contact with the mother's waste or infected contactants. The clinical signs  
185 in these pups develop faster and more severely because they still have an immature immune  
186 system. In adult animals, the disease usually has no symptoms (RODRIGUES & MENEZES,  
187 2003).

188 The parasite, when installed in the intestinal mucosa, causes ulcerations that can lead to  
189 blood loss and worsening of the disease, making room for secondary infections by opportunistic  
190 bacteria. It can also cause intestinal perforation, consequently triggering peritonitis and  
191 septicemia (TESSEROLLI et al., 2005).

192 Isosporosis is also a zoonosis, which usually affects immunosuppressed patients, the  
193 elderly and children. The observed clinical manifestations are directly linked to the immune  
194 system. Intense diarrhea and dehydration are the most common symptoms. Parasitosis may  
195 evolve into a chronic infestation, characterizing a syndrome of intestinal poor absorption  
196 (VASCONCELOS et al., 2008; PEREIRA et al., 2009).

197 The risk of infection in humans is high in the study area considering that the three species  
198 found have zoonotic potential, and in general sanitary education measures are necessary.  
199 Especially considering that pets and wandering animals contribute significantly to  
200 environmental contamination through their wastes deposited in urban areas (ENGBAEK et al.,  
201 1984).

202



203  
204  
205  
206  
207  
208  
209  
210  
211  
212  
213  
214  
215  
216  
217  
218  
219  
220  
221  
222  
223  
224  
225  
226  
227

## CONCLUSIONS

The occurrence of gastrointestinal parasites in animals, mainly domestic, is of great importance in both animal and human health. The findings in the research point to *Toxocara* spp. and *Ancylostoma* spp. as the most common species in the city of Mineiros, Goiás. The result presented occurrence of zoonotic species, showing the need to develop control programs to reduce the occurrence of helminths and protozoa in animals, since the presence of these parasites cause significant delay in their development, especially in young ages, and also exposure of these zoonotic agents to humans in Mineiros, Goiás, Brazil.

## REFERENCES

- BOWMAN, D. D. **Georgis' Parasitology for Veterinarians**. 10.ed. Saint Louis: Elsevier Saunders, 2014, p. 496.
- ARAÚJO, F. R.; CROCCI, A. J.; RODRIGUES, R. G. C. Contaminação de praças públicas de Campo Grande, Mato Grosso do Sul, Brasil, por ovos de *Toxocara* e *Ancylostoma* em fezes de cães. **Revista da Sociedade Brasileira de Medicina Tropical**, v. 35, n. 5, p. 581-583, 1999.
- BUSH, A. O.; LAFFERTY, K. D.; LOTZ, J. M.; SHOSTAK, A. W. Parasitology meets ecology on its own terms: Margolis et al. revisited. **The Journal of Parasitology**, v.83, n. 4, p. 575-583, 1997.
- CAPUANO, D. M.; ROCHA, G. M. Ocorrência de parasitas com potencial zoonótico em fezes de cães coletadas em áreas públicas do município de Ribeirão Preto, SP. **Revista Brasileira de Epidemiologia**, v. 9, n. 1, p. 81-86, 2006.

228

229 ENGBAEK, K.; MADSEN, H.; LARSEN, S. O. A suvey of helminths in stray cats from  
230 Copenhagen With ecological aspects. **Zeitschrift fur Parasitenkunde**, v. 70, n. 1, p. 87-94,  
231 1984.

232

233 HOFFMANN, R. P. **Diagnóstico de Parasitismo Veterinário**. Porto Alegre: Sulina, 1987, p.  
234 156.

235

236 LEIPER, R. T. Two new general of nematodes occasionally parasitic in man. **The British**  
237 **Medical Journal**, v. 1, p. 1296, 1907.

238

239 MARTINS, I. V. F. **Parasitologia Veterinária**. Vitória: Edufes, 2019, p. 320.

240

241 NUNES, C. M.; PENA, F. C.; NEGRELLI, G. B.; ANJO, C. G.S.; NAKANO, M. M.; STOBBE  
242 N. S. Ocorrência de larva migrans na areia de áreas de lazer das escolas municipais de ensino  
243 infantil, Araçatuba, SP, Brasil. **Revista Saúde Pública**, v. 34, n. 6, p. 656-658, 2000.

244

245 PAWLOWSKI, Z. Toxocariasis in humans: clinical expression and treatment dilemma.  
246 **Journal of Helminthology**, v. 75, n. 4, p. 299-305, 2001.

247

248 PEREIRA, A. D.; DAMIN, J.; LIMA, L. M.; ULIANO, R. F. *Isospora belli*: aspectos clínicos  
249 e diagnóstico laboratorial. **Revista Brasileira de Análises Clínicas**, v. 41, n. 4, p. 283-286,  
250 2009.

251

252 RAMÍREZ-BARRIOS, R. A; BARBOZA-MENA, G.; MUÑOZ, J.; ANGULO-CUBILLÁN,  
253 F; HERNÁNDEZ E.; GONZÁLEZ, F.; ESCALONA, F. Prevalence of intestinal parasites in  
254 dogs under veterinary care in Maracaibo, Venezuela. **Veterinary Parasitology**, v. 121, n. 1-2,  
255 p. 11-20, 2004.

256

257 RAMOS, D. G. S.; SCHEREMETA, R. G. A. C.; OLIVEIRA, A. C.S.; SINKOC, A. L.;  
258 PACHECO, R. C. Survey of helminth parasites of cats from the metropolitan area of Cuiabá,  
259 Mato Grosso, Brazil. **Revista Brasileira de Parasitologia Veterinária**, v. 22, n. 2, p. 201-206,  
260 2013.

261

262 RAMOS, D. G. S.; ZOCCO, B. K. A.; TORRES, M. M.; BRAGA, I. A.; PACHECO, R. C.;  
263 SINKOC, A. L. Helminths parasites of stray dogs (*Canis lupus familiaris*) from Cuiabá,  
264 Midwestern of Brazil. **Semina: Ciências Agrárias**, v. 36, n. 2, p. 889-894, 2015.

265

266 RIBEIRO, V. M. Controle de helmintos de cães e gatos. **Revista Brasileira de Parasitologia**  
267 **Veterinária**, v. 13, suppl. 1, p. 88-95, 2004.

268

269 RODRIGUES, A. N.; MENEZES, R. C. A. A. Infecção natural de cães por espécies do gênero  
270 *Cystoisospora* (Apicomplexa: Cystoisosporinae) em dois sistemas de criação. **Clínica**  
271 **Veterinária**, v. 42, n. 1, p. 24-30, 2003.

272

273 SANTARÉM, V. A.; GIUFFRIDA, R.; ZANIN, G. A. *Larva migrans* cutânea: ocorrência de  
274 casos humanos e identificação de larvas de *Ancylostoma* spp. em parque público do município  
275 de Taciba, São Paulo. **Revista da Sociedade Brasileira de Medicina Tropical**, v. 37, n. 2, p.  
276 179-181, 2004.

277

278 SHARIF, M.; NASROLAHEI, M.; ZIAPOUR, S. P.; GHOLAMI, S.; ZIAEI, H.; DARYANI,  
279 A.; KHALILIAN, A. *Toxocara cati* infections in stray cats in northern Iran. **Journal of**  
280 **Helminthology**, v. 8, n. 1, p. 63-66, 2007.

281

282 SHERDING, R.; JOHNSON, S. Doenças intestinais. In: BIRCHARD, S. J.; SHERDING, R.  
283 G. (Eds.) **Manual Saunders - Clínica de pequenos animais**. 3.ed. São Paulo: Roca, 2008, p.  
284 719-756.

285

286 TAYLOR, M. A.; COOP, R. L.; WALL, R. L. **Veterinary Parasitology**. 3.ed. Oxford:  
287 Blackwell Publishing, 2007, p. 874.

288

289 TESSEROLLI, G. L.; FAYZANO, L.; AGOTTANI, J. V. B. Ocorrência de parasitas  
290 gastrointestinais em fezes de cães e gatos, Curitiba – PR. **Revista Acadêmica de Ciências**  
291 **Agrárias e Ambientais**, v. 3, n. 4, p. 31-34, 2005.

292

293 VASCONCELOS, M. G. C.; TALON, D. D. B.; SILVA-JÚNIOR, C. A.; NEVES, M. A.;  
294 SACCO, S. R. Isosporose nos animais domésticos. **Revista Científica Eletrônica de Medicina**  
295 **Veterinária**, v. 6, n. 10, p. 1-7, 2008.

296

297 WILLIS, H. H. A simple levitation method for the detection of hookworm ova. **The Medical**  
298 **Journal of Australia**, v. 8, p. 375-376, 1921.

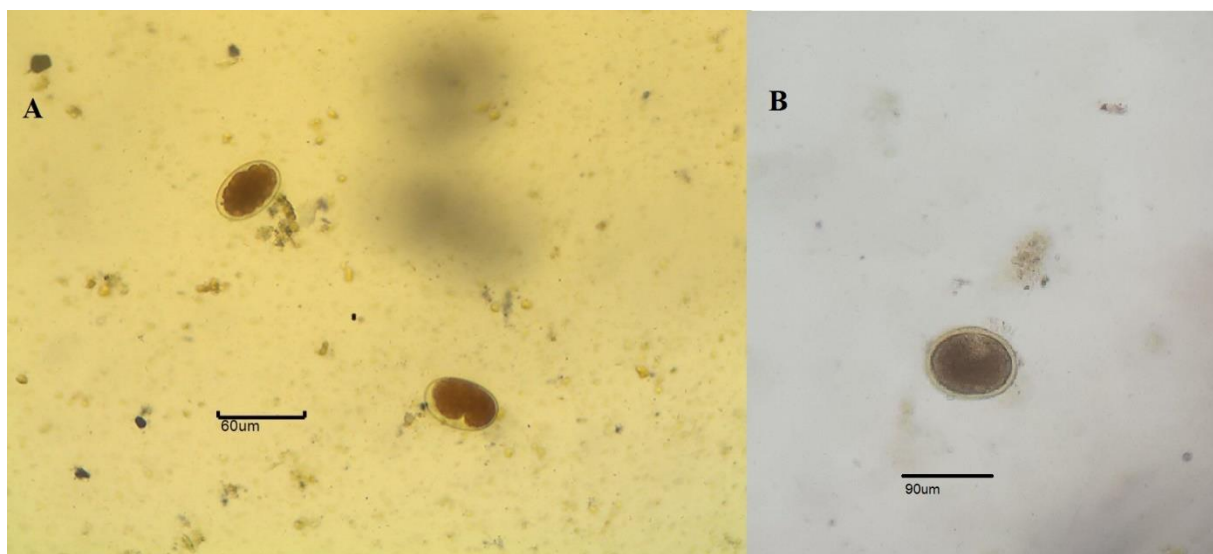
299

300 WOODRUFF, A. W.; SALIH, S. Y.; SAVIGNY, D.; BAYA, E. I., SHAH, A. I.; DAFALLA,  
301 A. A. Toxocariasis in the Sudan. **Annals of Tropical Medicine and Parasitology**, v. 75, n. 5,  
302 p. 559-561, 1981.

303

304 ZAJAC, A. M.; CONBOY, G. A. **Veterinary Clinical Parasitology**. 8.ed. Oxford: Wiley-  
305 Blackwell, 2012, p. 368.

306



307

308 Figure 1. Helminth eggs identified in fecal samples of dogs and cats from Mineiros, Brazil,  
309 from April 2017 to July 2018 under optical microscopy (scale bar in figure). A) Eggs of  
310 *Ancylostoma* spp.; B) Egg of *Toxocara* spp.